

TMM 3102: Protein Structure, Function and Disease

- Structural Biology Methods: Molecular Dynamics Simulation
(October 7th, 2021)

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(Partially adopted from former lectures by Dr. Maria Musgaard)



Importance: Static v.s. Dynamic



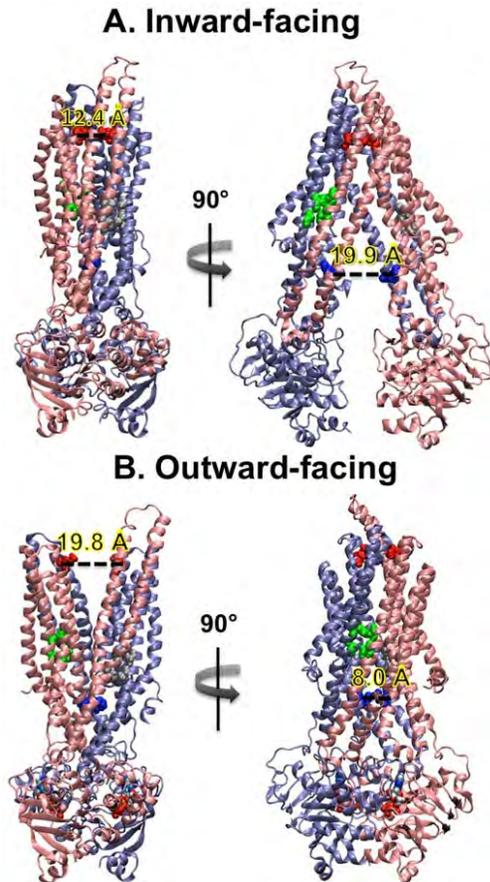
DYNAMIC



STATIC

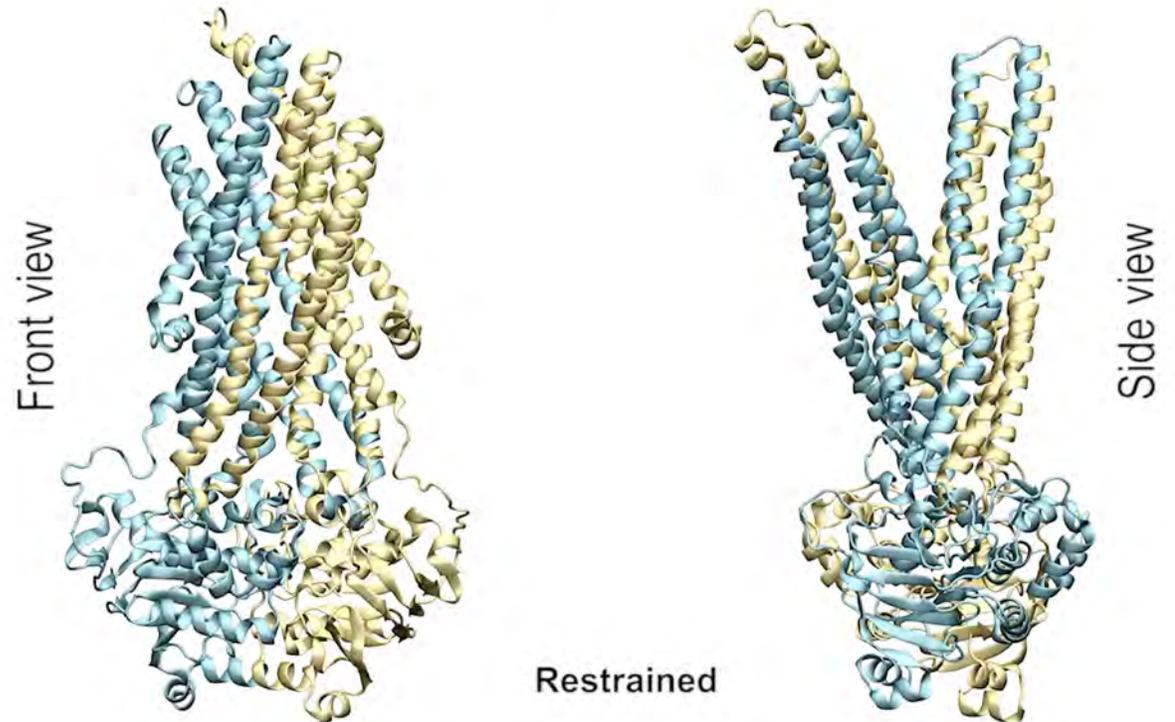
(<https://digitos.io/benefits-of-dynamic-digital-signage-over-static-signage/>)

Importance: Static v.s. Dynamic



(Pan & Aller, *Sci Rep*, 2015)

Dynamics of outward-facing (OF) state of Pgp in membrane



(Verhalen et al, *Nature*, 2017)

Bridging the Gap

Protein function

- Functional data
- Electrophysiology
- Substrate transport
- ...
- High resolution in time

Protein structure

- X-ray
- NMR
- Cryo-EM
- ~ "snapshots"
- High resolution in space

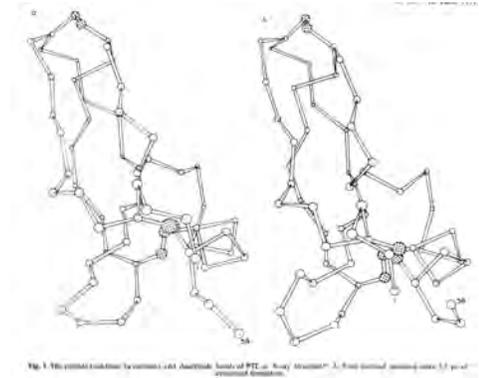


High resolution in
"space" and "time"?

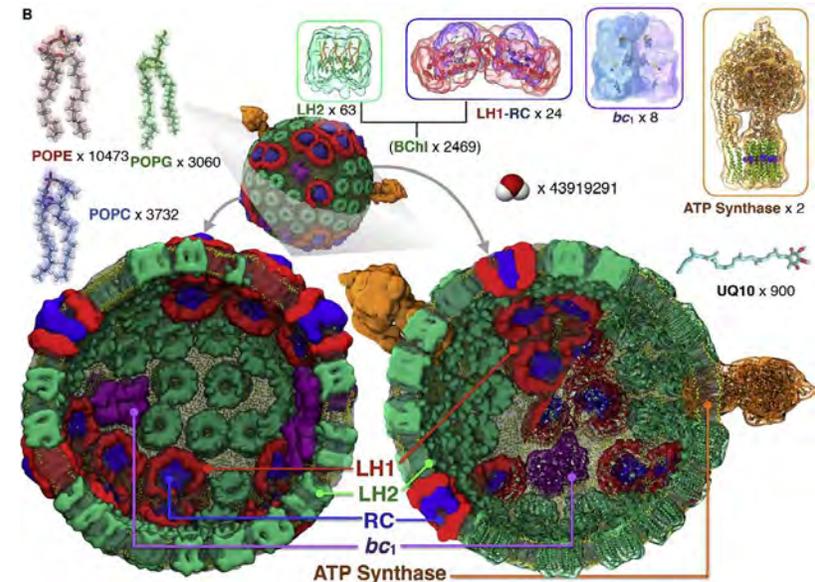
Brief History

- First MD study of proteins published in 1977
~60 residues, no solvent, ~9 ps
- 2019: full organelles, 139 million atoms, 0.5 μ s

- Factors:
 - more structures determined
 - better algorithms
 - faster computer



(Nature, 1977)



(Cell, 2019)

Molecular Dynamics (MD): idea

- Classic mechanics (thinking of “Newton’s laws of motion”)
- Metaphor:
If cycling at 15 km/h by Canal Rideau; keep a constant acceleration:
 - Predict how long to reach uOttawa main campus.
 - Predict where you are in 5 minutes.
- Do the same for all atoms in a protein system

Molecular Dynamics (MD): idea

Going to the next position:

- $r(t+\Delta t) = r(t) + \Delta t*v(t) + 1/2[\Delta t^2*a(t)]$

$r(t)$: position at "t"

$r(t+\Delta t)$: position after Δt

$v(t)$: velocity

$a(t)$: acceleration

Molecular Dynamics (MD): idea

Acceleration:

$$F = m * a$$

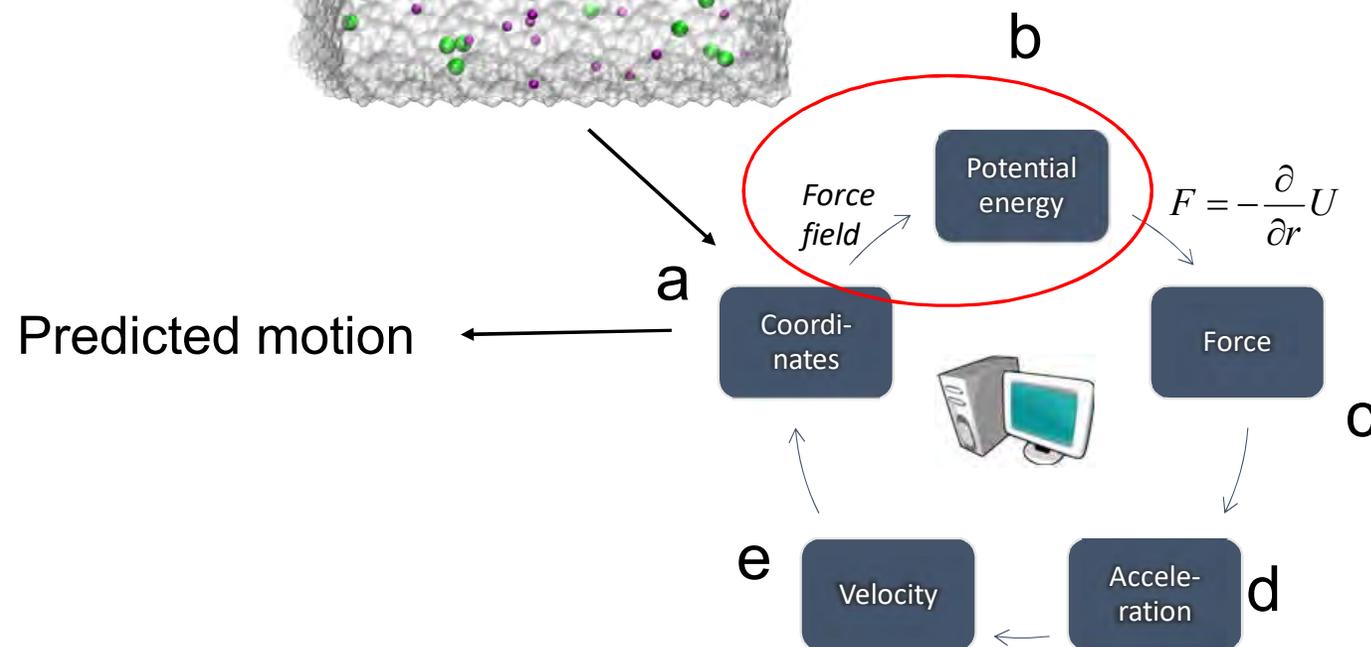
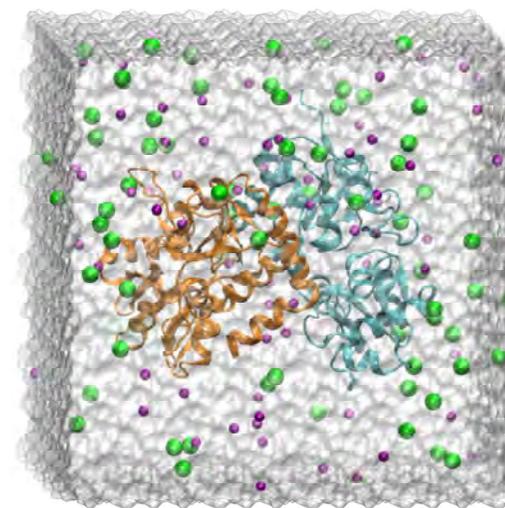
$$F = -\Delta U / \Delta r$$

If we know U (potential energy), then we can calculate the force and the acceleration on each atom.

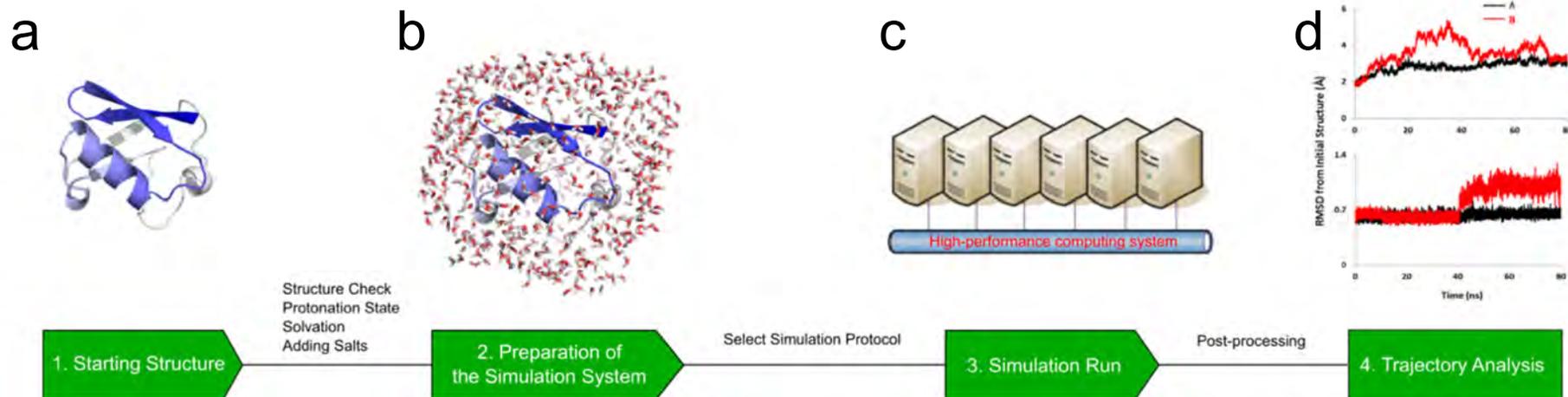
Molecular Dynamics (MD): workflow

In general, how do we do MD simulation?

- Find the coordinates of a known protein model from the database.
- Choose a force field to generate energy potential for further calculation.
- Calculate the force that results from the theoretical potential energy.
- Find out how molecules speed up with the obtained force.
- Calculate the speed of the molecule and where the protein move into.



Molecular Dynamics (MD): workflow



Another way to see the MD workflow:

- Find a model template and artificially add necessary ingredients that suit the physiological condition of the target protein. This includes protonation states, salts, water, etc.
- Prepare the simulation system by selecting the best protocol, aka force field.
- Run the simulation using a cluster of computers.
- Process the data and predict the where the segment of interest moves to.

Molecular Dynamics (MD): force field

What determines “force field”?

- Atoms: different in size, softness, mass, charge, ...
- Bonds: different in lengths, stiffness, ...
- Electrons: implicitly accounted for covalent bonds.

Molecular Dynamics (MD): force field

What is a force field used for?

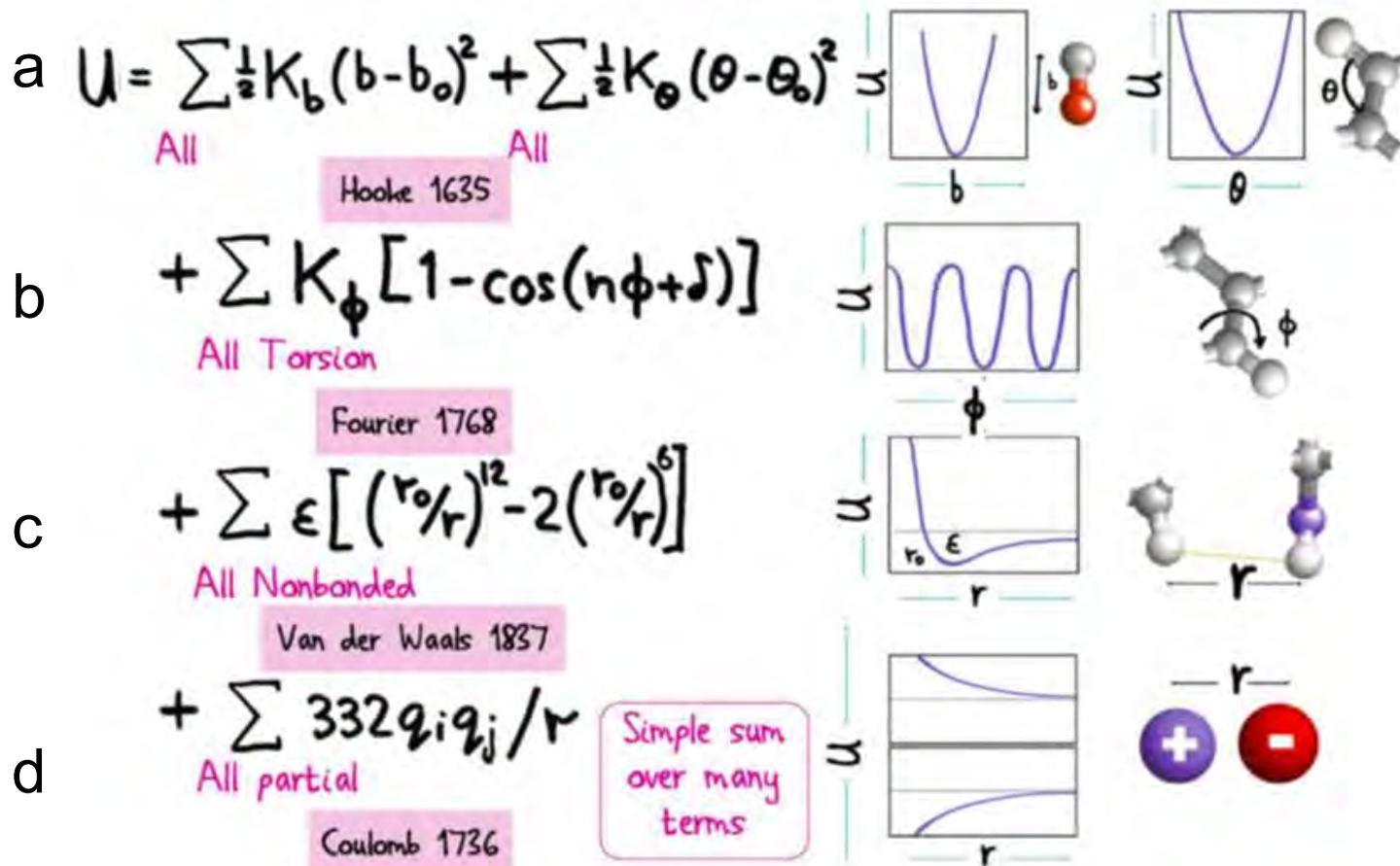
- Used for large molecules or conformational studies
- Not used to break or form chemical bonds
- Empirical, so no one is most correct.
- Requires:
 - Energy equation to describe U as a function of atomic coordinates
 - Constant parameters to be used in the energy equation
 - Atom types to establish constant parameters, charges, masses, etc.

Molecular Dynamics (MD): force field

MOLECULAR POTENTIAL ENERGY

Selection of force field is like deciding what kind of potential energy to use:

- Covalent bonds & bond angles
- Torsion angles
- Van der Waals interaction
- Electrostatic force / charge-charge interaction

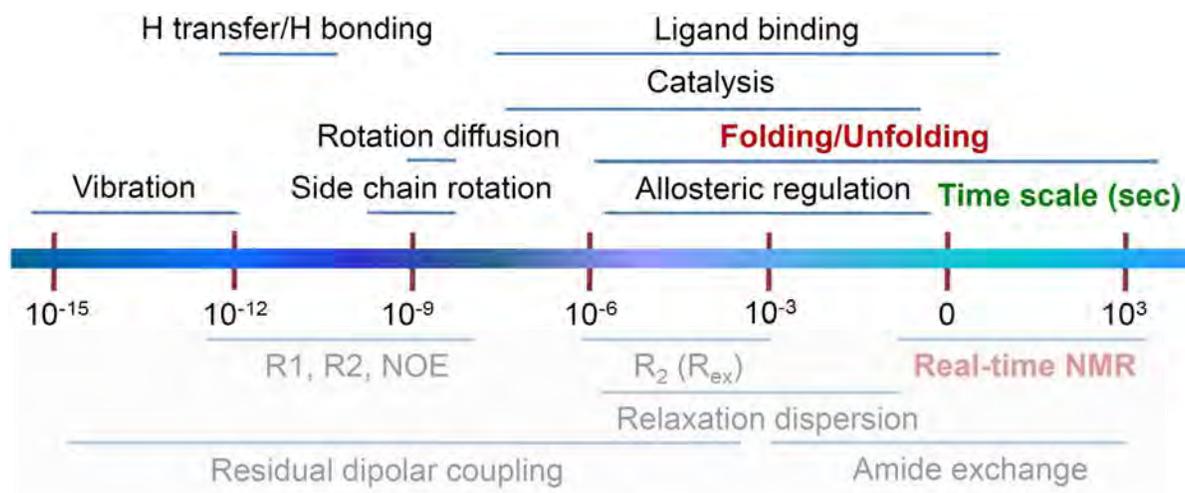


Molecular Dynamics (MD): force field

- Do's and Don'ts
 - Never compare energies from different force fields, unless absolute energy is known
 - Never mix parameters, unless tested
 - Do simulations in the conditions similar to those used to obtain the force field
 - For new ligands, need a full set of parameters (all you can)

Molecular Dynamics (MD): time scale

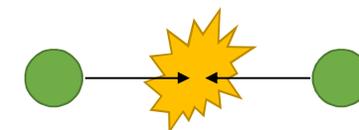
Biological timescales



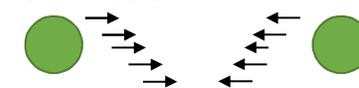
Kumar and Balbach, *Biochim. Biophys. Acta* 2015

- Simulation Δt : 1-2 fs

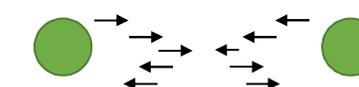
- Too fast:



- Too slow:

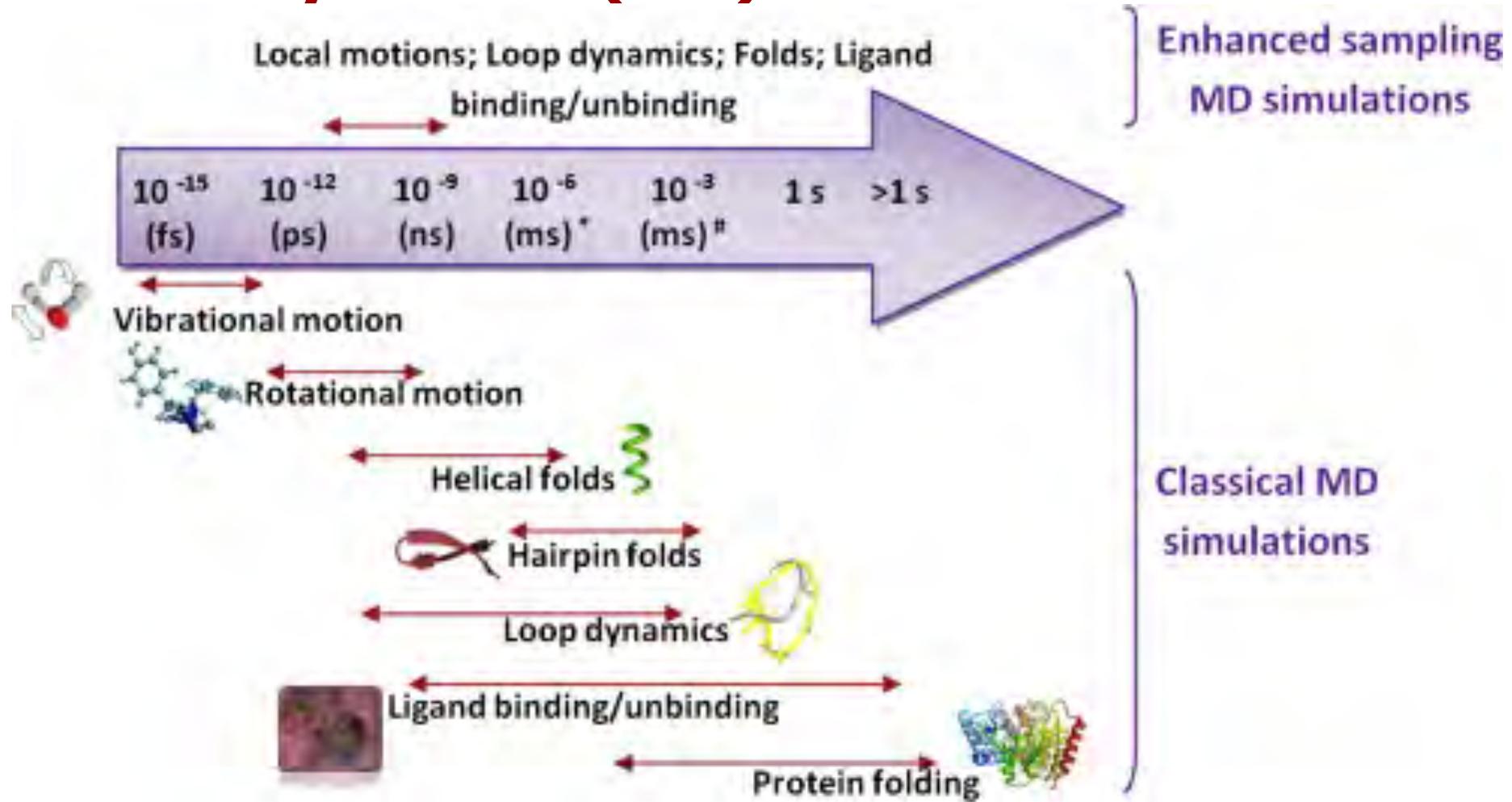


- Good



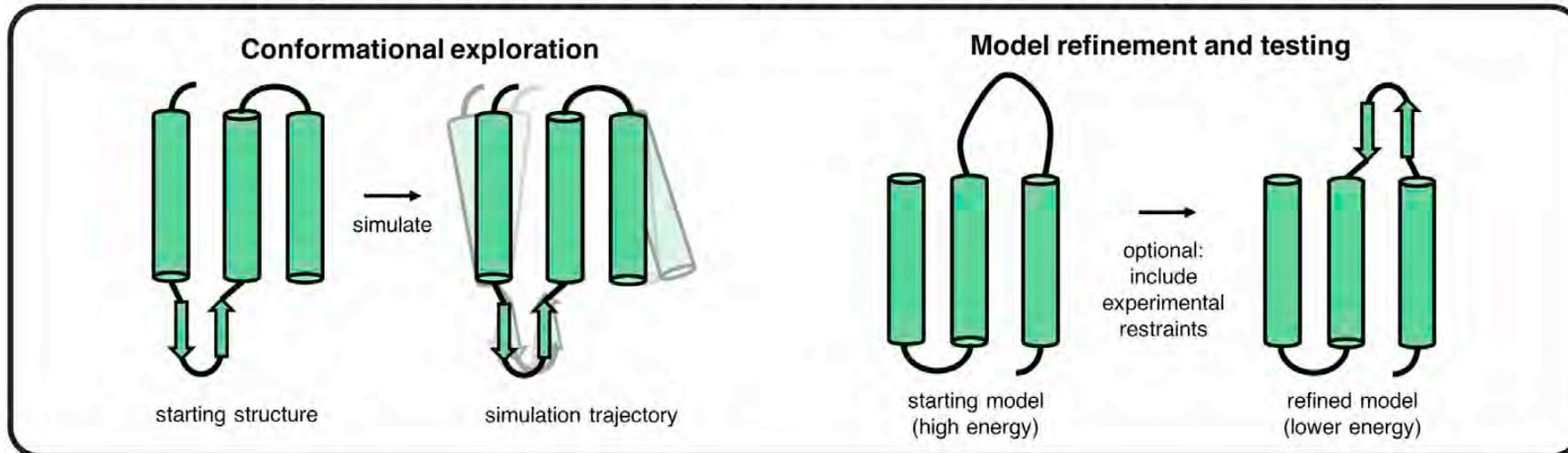
- => 0.5 to 1 million steps to reach 1 ns (!)

Molecular Dynamics (MD)



Structural Determination *in silico*

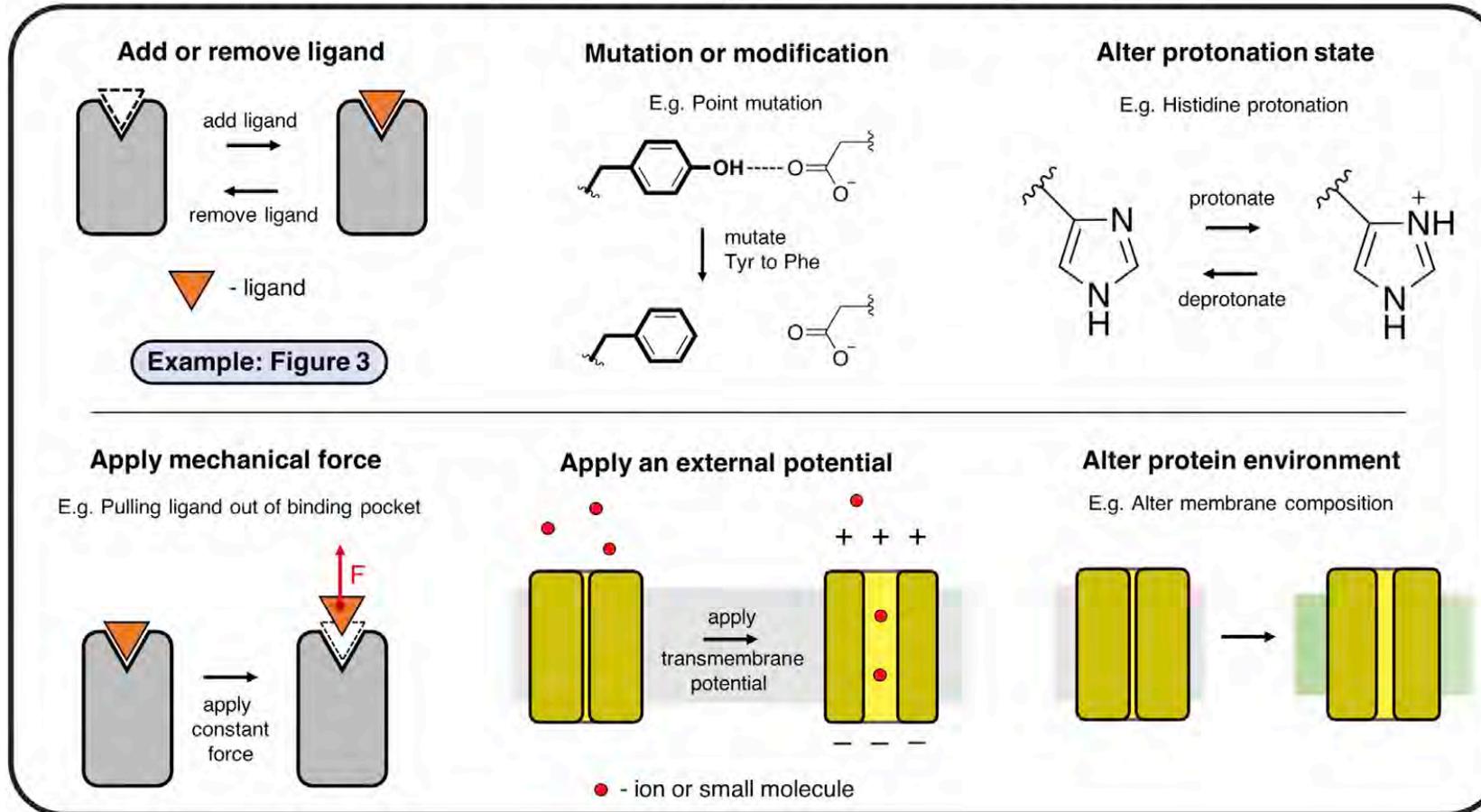
Structural and dynamic studies: *Studying conformational flexibility and stability*



(Hollingsworth & Dror, *Neuron*, 2018)

Structural Determination *in silico*

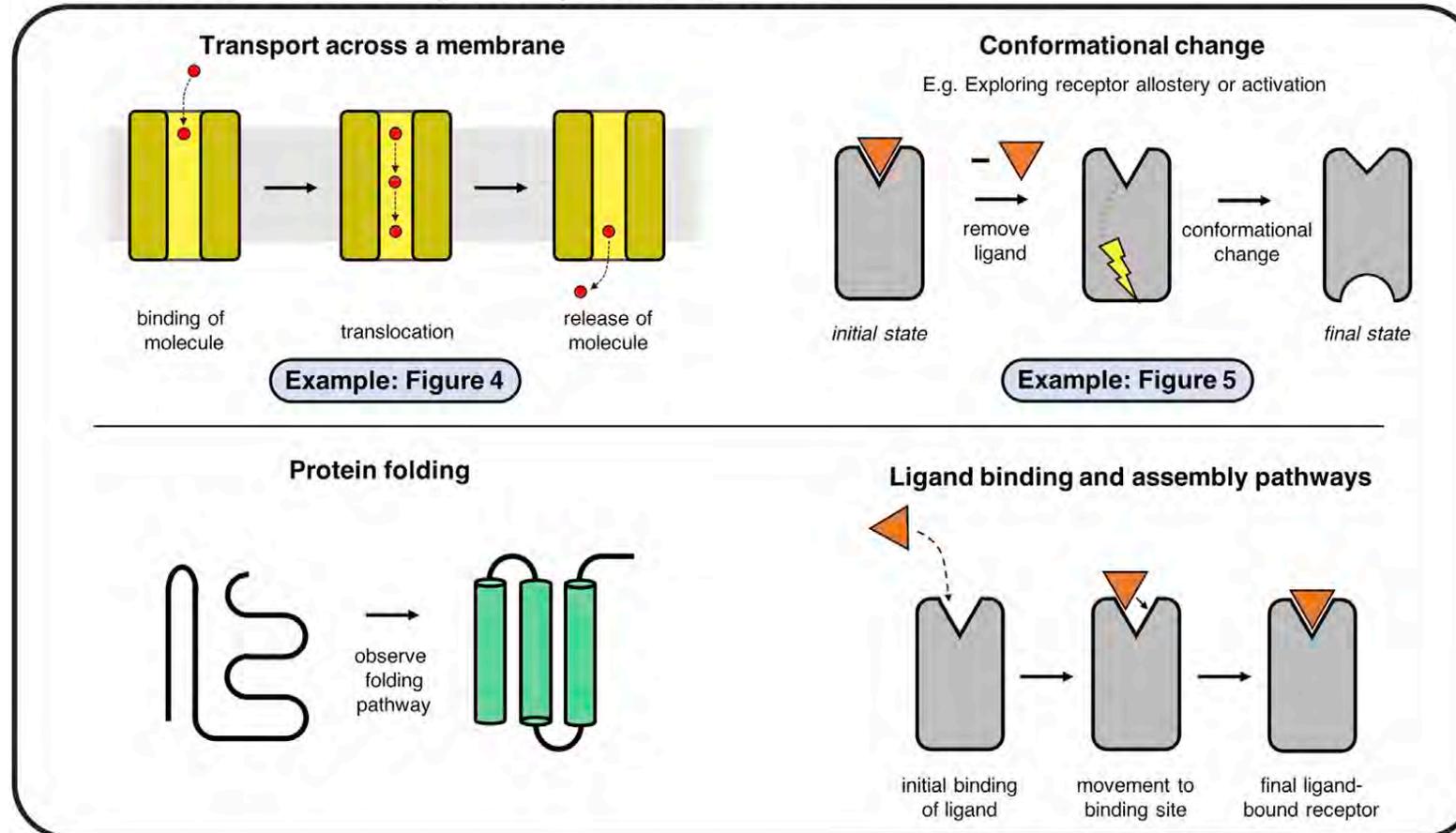
Perturbations: Observe response following controlled change to system



(Hollingsworth & Dror, Neuron, 2018)

Structural Determination *in silico*

Processes: *Observe a dynamic process over time*

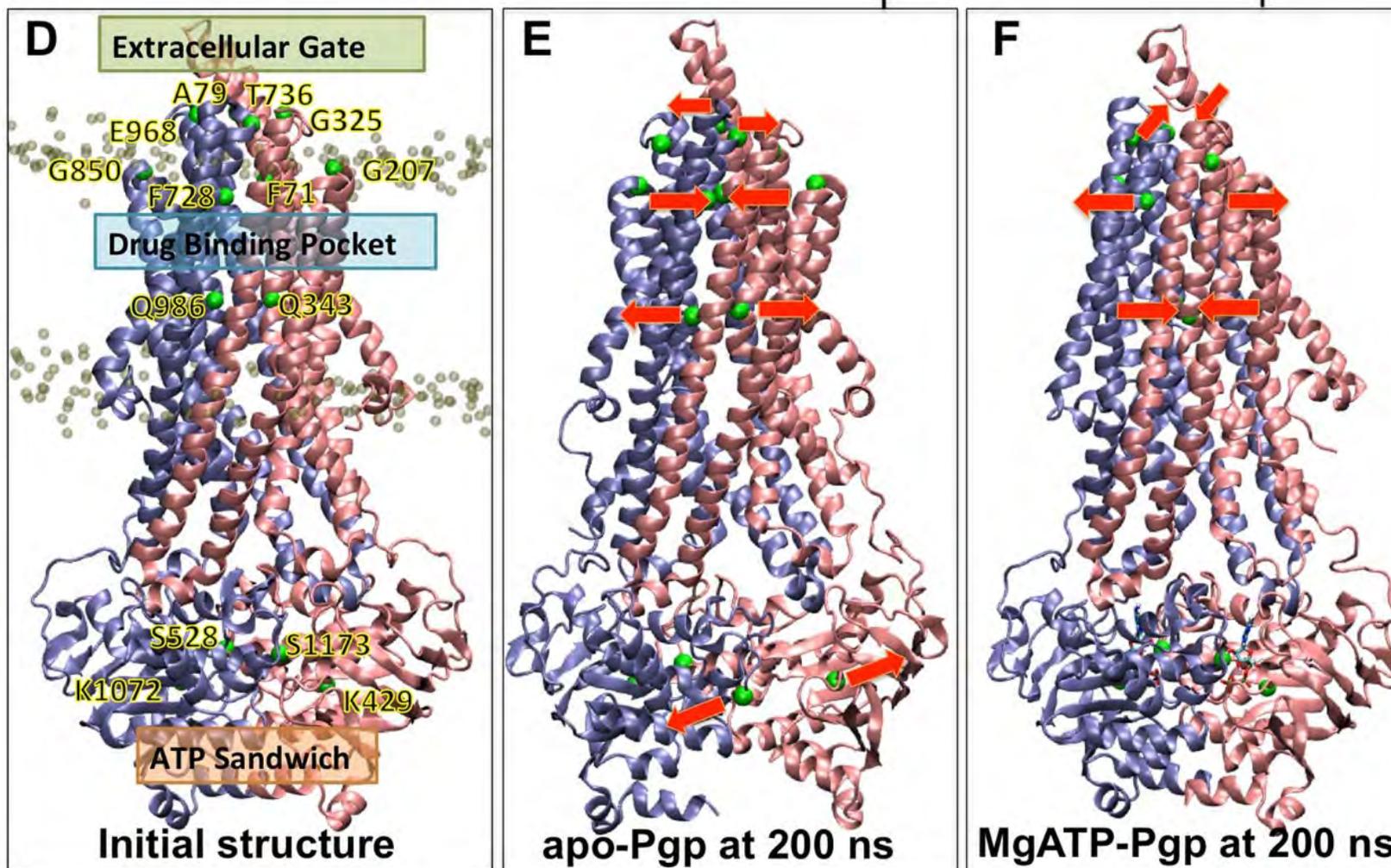


(Hollingsworth & Dror, Neuron, 2018)

Molecular Dynamics (MD)

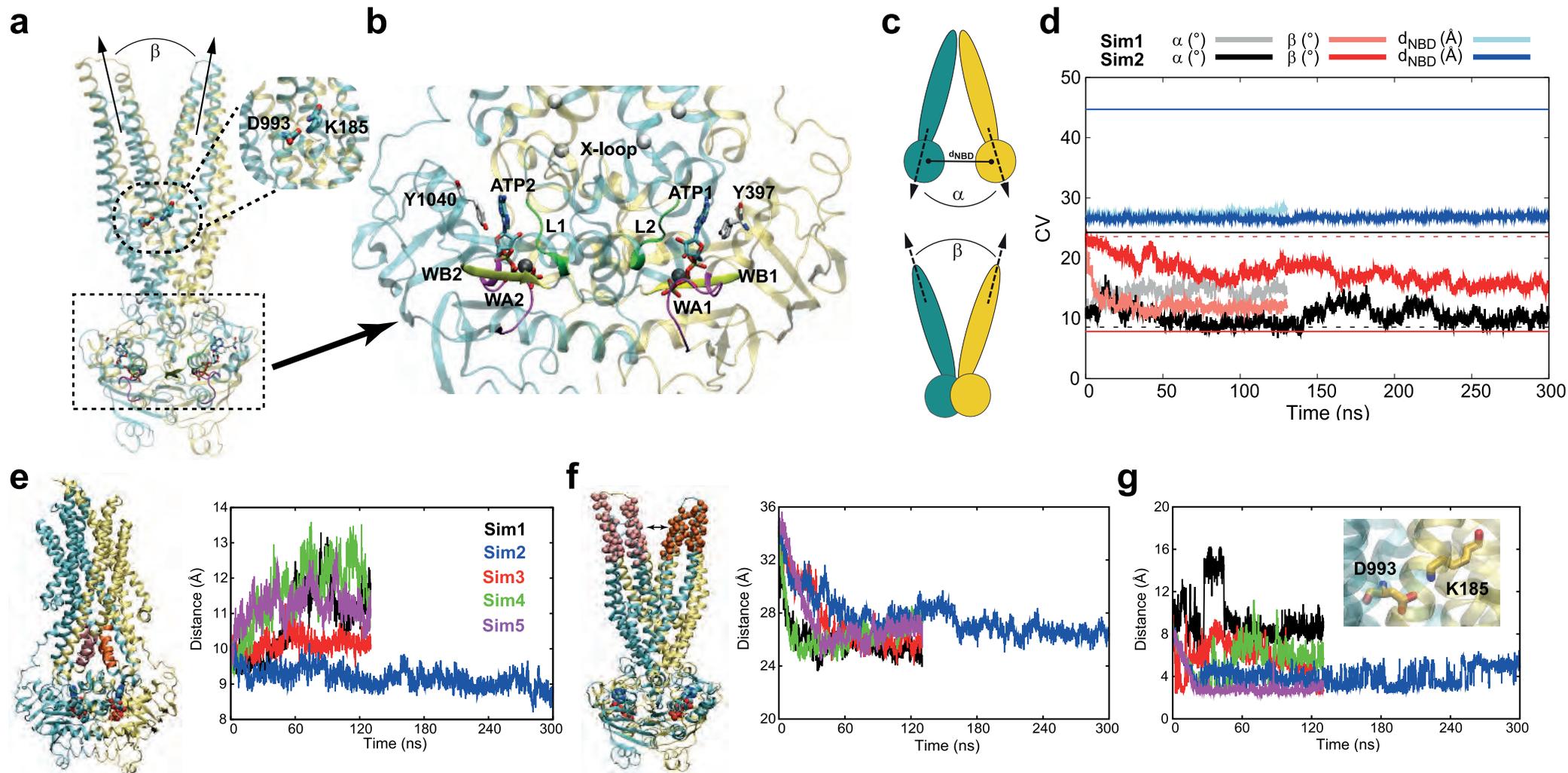
- Advantages
 - High resolution in space and time
 - Precise simulation conditions: conformations, \pm ligands, ...
 - Cheap: mutations, protein-ligand, protein design, ...
 - Structure-function relationship
- Limitations
 - Validation: need experimental data
 - Timescale and sampling
 - Quality of starting structures
 - Force fields
 - No bond making/breaking, as it depends on protonation states

Case Study: P-glycoprotein (drug-resistance)



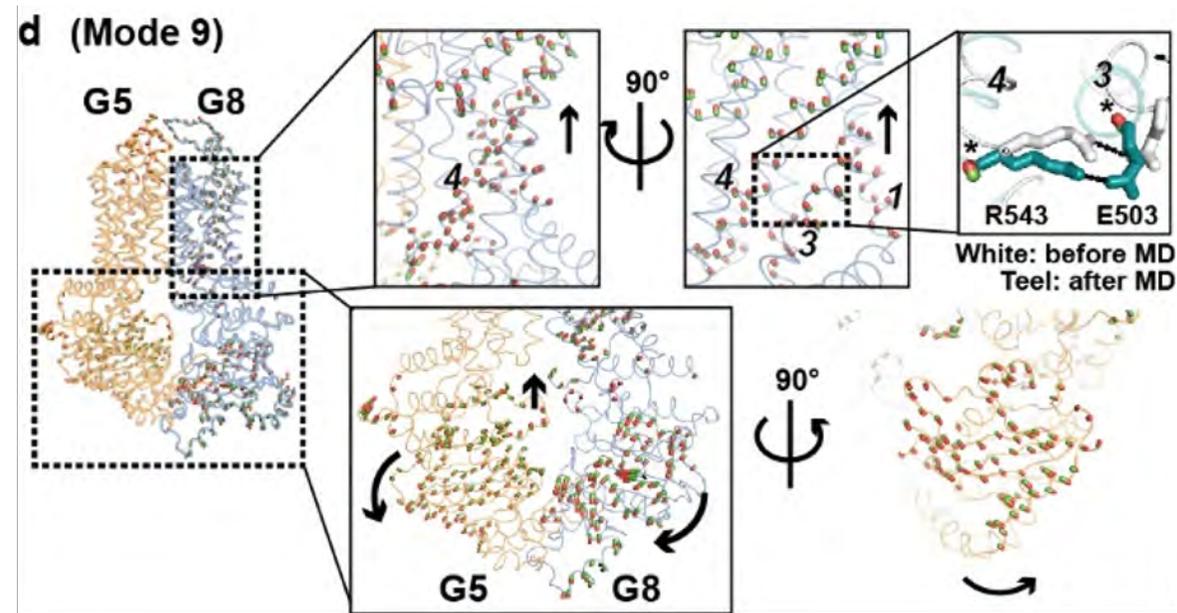
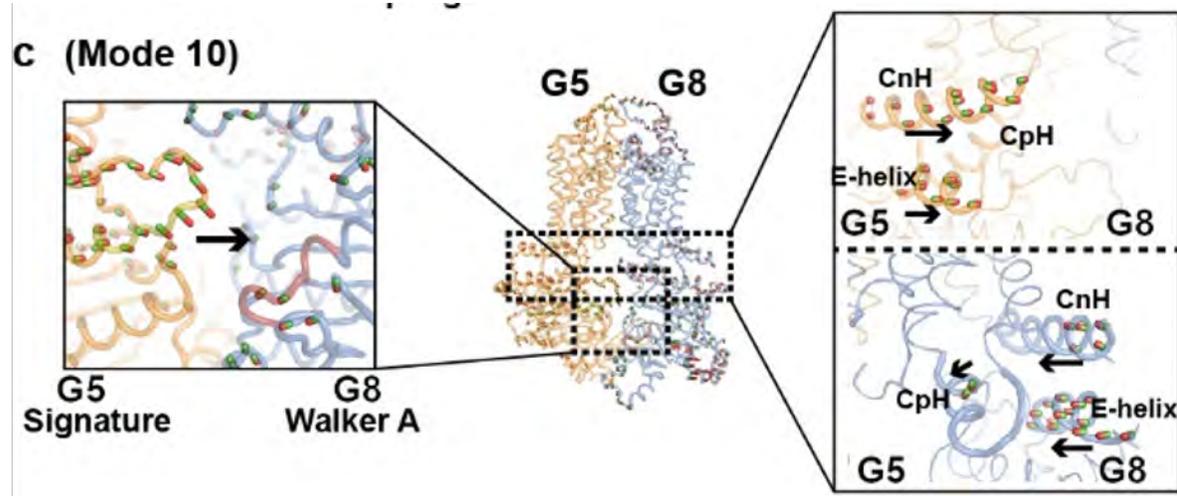
(Pan & Aller, *Sci Rep*, 2015)

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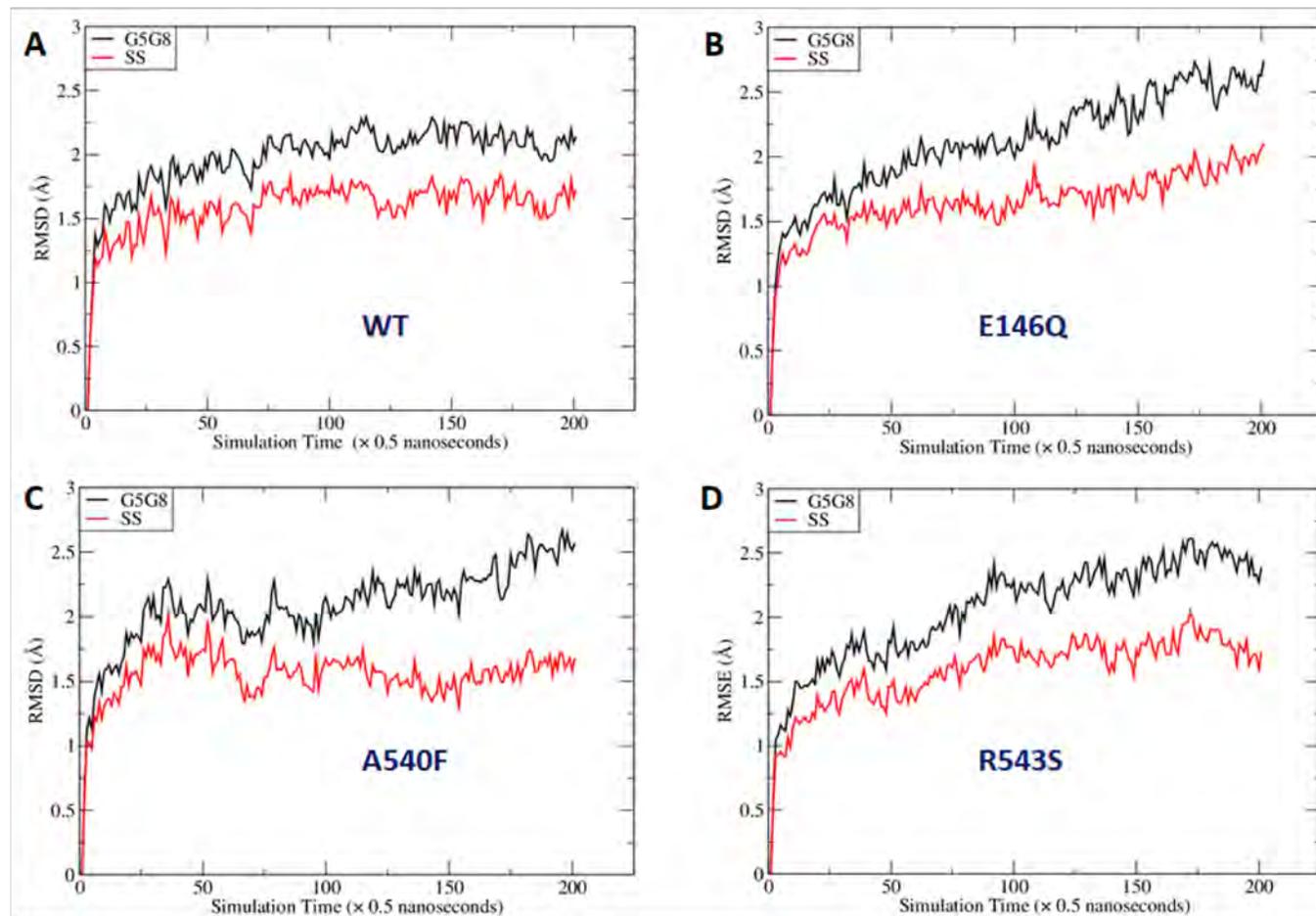
(Verhalen et al, Nature, 2017)

Case Study: ABCG5/G8 (sterol efflux)



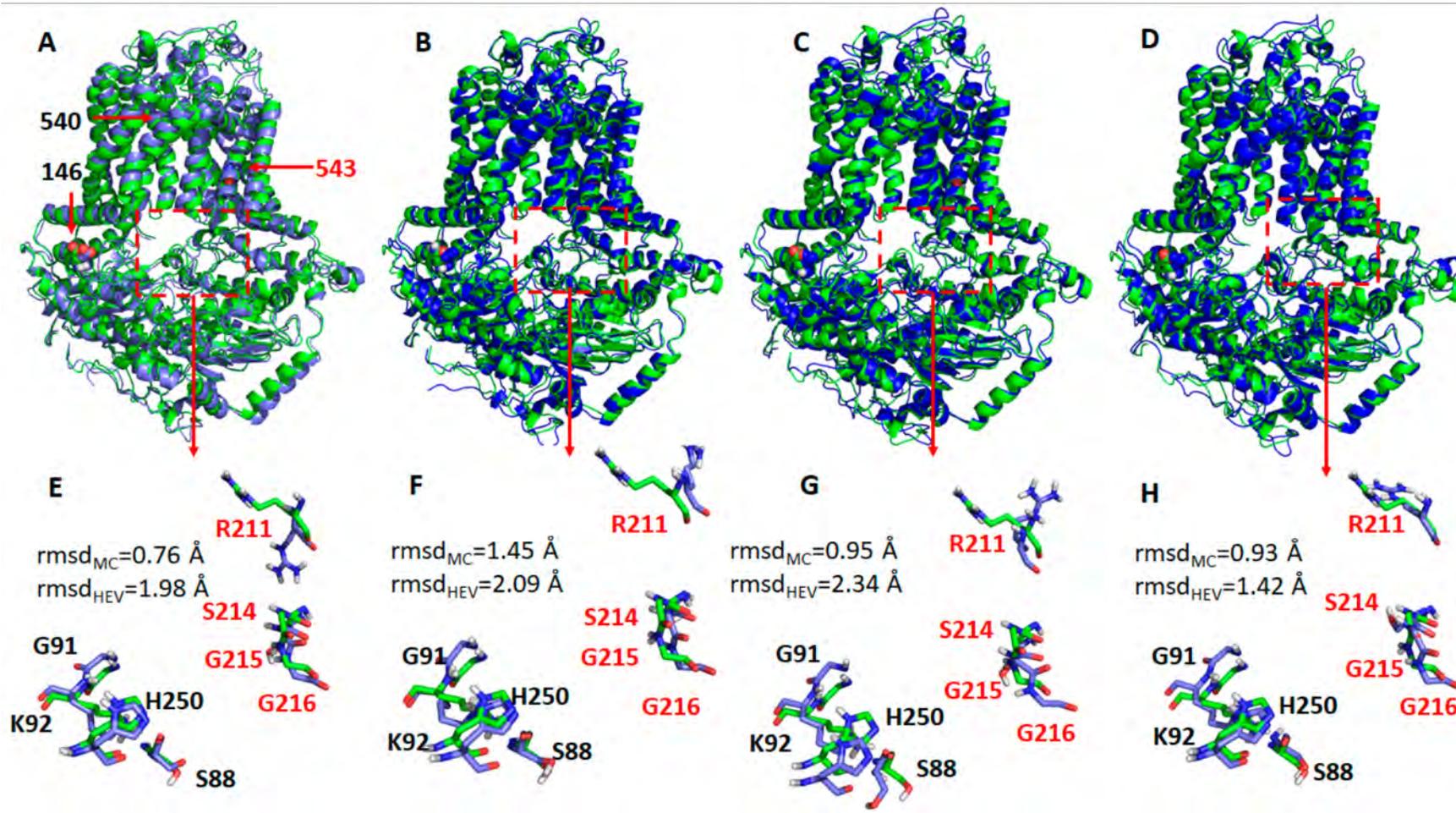
(Lee et al, Nature, 2016)

Case Study: ABCG5/G8 (sterol efflux)



(Xavier et al, IJMS, 2020)

Case Study: ABCG5/G8 (sterol efflux)



(Xavier et al, IJMS, 2020)